

Coment ID	Commen t Text	Comment Response	Page location of primary response	EPA QA RESPONSE - J.Crawford 9-8-11	Additional clarification requested
VAC1	Gina is actually the RQAM. The QA Chemists reviewing QAPPs have delegated authority to approve the plan for her, so I sign in her place.	Noted and included in revision, see pg	1	Complete	
VAC2	Please reference the WCD QAPP here with a citation as well. (I do see one below in 1.1)	Noted and included in revision, see pg	4	Complete (1.0)	
VAC3	Are matrix spikes going to be conducted at a rate of 5% for the project (similar to the WCD project)?	Groundwater samples are expected to be dilute and not likely subject to significant matrix effects during annalysis. However, this will be testeds during the first sampling event specifcally in samples with large specific conductance values.	13	Complete (3.0)	
NU4	Also, what is the frequency for these replicate and duplicate samples? 5%?	About 5 % each. Total of all QA samples from the field will be about 15-20%	14	Noted, please state frequency goal in QAPP 3.0 Precision Section	Each sampling event will include at least 1 field blank and 1 replicate sample per every 20 samples submitted to the lab
NU5	Relative Percent Difference is a measure of precision (see above). How about "percent recovery" instead? And need to provide the formula for it's calculation like was done for RPD under the precision section.	RPD is planned as a measure of accuracy when applied to a reference sample. When matrix spikes are added to check for matrix interference, percent recovery will be used as a measure of accuracy. Formula added to text.	15	Complete (3.0 Accuracy)	
VAC6	Great! What criteria will there be for these splits?	If analytical results from sample splits exceed two times the field replicate samples the source of the variability will be investigated. It should be noted that USGS and WCD project chiefs anticipate having detailed discussions very early in the sampling process to optimize SOPs so that comparability of the data generated is at the highest practical level.	17	Complete (3.0 Comparability)	
VAC7	This only covers one part of the QC involved. Lab analyses should have their own QC table identifying the Measurement Quality Objectives for QC. This should be parsed out by each individual analysis to mirror Table 4.	Table modified. Laboratory control limits are based on the f-psuedosigma meausre of the data generated from control samples which including blanks, continuing calibration standards, and third party reference standards. Dispersion of the measured values of the control samples from the expected concentrations is expressed using the f-psuedosigma, equivalent to the standard deviation divided by 1.349. See Helsel nd Hirsch. Statisitcal Methods in Water Reosurces. When continuing control calibration measurements are outside of the control limits, affected analysis are rurun.	18	Complete (Table 3) - would prefer a numeric 'goal' criteria so there is an idea of expected accuracy, but the statistical criteria applied by the lab is likely more stringent than a standard method specified range.	Table 3 does give an expected accuracy ot about 25 %, but the lab control limits are typically more stringent.
IC8	Verifying method w/ EPA microbiologists to ensure comparability to other WCD analyses	Noted, see comments below labeled micro1-micro6			

on page 14

JC9	Are the methods listed the current methods NWQL is performing? Or can they be updated to match the EPA approved methods listed in the 2007 Methods Update Rule? Although not a requirement (no regulatory requirement here) it is always recommended. Overall I want to get as comparable laboratory data as possible for the USGS and WCD data. I noted the method listed in MUR for reference. It is also stated in the comparability section that they will use 40 CFR 136 (i.e., MUR) comparable methods.	Methods listed are current with NWQL. There maybe an issue as NWQL transitions colorimetric nitrate reduction analysis from cadmium reduction to nitrate-reductase method.	25	Note the switch to alternate nitrate reduction in the future, perhaps as a footnote to Table 4. Will this be covered by a separate USGS method?		footnoted in table 4
VAC10	Since everything is field filtered, analyses would be more accurately labeled 'dissolved' for clarity.	Using a .45 micron filter is an operational definition of 'dissolved' and should be distinguished from conditions when ions are simply hydrated and truly dissolved..		True. However, from an analytical perspective this is standard and consistent terminology for reporting the filtered water matrix. The USGS methods listed all include 'dissolved' for the matrix in the title/description. As long as the final results are reported as filtered/dissolved samples then I am satisfied with the Table 4 and 5 as noted.		OK, no real issues here, will report every thing as dissolved.
VAC11	Field preserved H2SO4	Acid preservation not required for short, chilled, darkened hold times. See results of QA demonstrations study showing that when biota are removed from samples at collection sites by 0.45-micrometer membrane filtration, subsequent preservation with sulfuric acid or mercury (II) provides no statistically significant improvement in nutrient concentration stability during storage at 4 degrees Celsius for 30 days. Patton and Gilroy 1999, US Geological Survey nutrient preservation experiment : experimental design, statistical analysis, and interpretation of analytical results: USGS WRIR 98-4118	28	Complete (Table 4) - please remove the method comments I added to the 'Method Number' column. Assume 'short' is defined as 30 days. This is acceptable. I am concerned with the number of analytical differences between WCD and USGS samples, but the split samples will speak to the comparability of the data sets. There is an inherent amount of variability already with the different methods and labs, so the altered preservation/matrix (total vs filtered) will just be one more layer. (Hopefully not much, according to the USGS publication cited.)	A couple additional questions: (1)Chloride HT listed: 180 is much longer than the standard EPA 28 days - is 180 days the std USGS HT? That is usually just for metals for us usually. (2) The NO3+NO2 method lists the analytical range lowest std as 0.1 with the applicable range starting at 0.05 - do they report all the way below this to 0.002? Are there any check standards lower? 2ppb is very low, so I am curious. (3) should E.coli be listed under lab instead of field?	1) 180 day HT changed to 28 day. Will specify to lab on sample submission the shorter hold time limitations required by project. 2) the lab reports estimated values below the quantitation limits, all estimated values are noted and can be censored. 3)E. coli is a field measurement, Any lab measurements are part of QA
JC12	DA = ?	typo		DA is still listed in Table 4		
VAC13	Field preserved ZSO4	Acid preservation not required for short, chilled, darkened hold times, see above comment VAC11	28	Same comment as above. Complete (Table 4).		
JC14	Are potassium and iron being analyzed by difference ICP-AES methods?	yes, different ICP method numbers for cations and metals	25	I looked up the USGS method and the Potassium is actually a Flame AA analysis, not ICPAES from what I can tell.	Flame is an old method, (lab needs to update webpage) ICP-AES method used for K is Standard Method 3120. Table 4 updated	
JC15	Section 4.6.1.2 also lists Total Phosphorus as an analysis. Add to table if correct.	noted and modified	26	Complete (Table 4)		
VAC16	Missing RUC code (E.coli)	Noted and inserted designation for bacteria samples	27	Added RUC, but is listed as RUC-Ster.	Updated RUC-Ster	

JC17	Preservation of nutrient samples with H2SO4 in field at collection for analysis by colorimetric methods is usually required – EPA MUR 2007, 40 CFR 122/136	Acid preservation will disrupt the analysis method used in the NWQL colorimetric determination. If acid preservation is required then a different laboratory will be needed. Additional acid preserved splits can be added to sampling plan and sent to accredited lab as check on sample degradation.	28	Complete (table 4) - comment above	
JC17	http://www.epa.gov/fedrgstr/EPA-WATER/2007/March/Day-12/w1073.pdf . If this is not standard USGS protocol, could it be done for better comparability to WCD sample data?	Comparability with WCD data will be assessed. Discussions of comparability	17	Complete, covered in various sections of QAPP. Concerns noted above but assessed with QA/QC samples and data sharing planned for project with WCD.	
JC18	Figure 3 instead?	Wrong figure number noted and corrected	22	Complete (4.4.3)	
VAC19	The chain of custody form does not include a section for transference of custody.	Sample shipment is handled under FedEx Shipping Airbill which are signed upon shipping and receipt. Once received by the lab the Login process opens the cooler measures and records the temperature of the contents of the cool using an infrared detector. the record of the receipt, temp, and initials of the person receiving the cooler are recorded on the ASR, a pdf record is attached to the sampleID record and the information is also recorded on the Laboratory information system. see Maloney 2005 for more details	30	Please note at the end of section 4.4.3 that the Airbill will be used as the custody transfer as stated in your comment. (I don't see this updated)	done
JC20	Recommend adding a column for the detection limit (sensitivity) of the instruments, or the calibration ranges.	Column added	31	Complete (Table 6)	
JC21	Is each sampling event more than one day? Recommend also checking the equipment at the end of each sampling day to verify the parameters are still calibrated and all data logged for the day is valid.	This is done. Text indicates that at the end of the sampling day another cal check is performed to check for monitoring instruments for drift.	31	Complete (4.5.1)	
VAC22	This is not the method referenced in table 4 (1-1472-87)	USGS analysis method identification for analysis of iron checked on table 4 and text.	34	I still see method I-4471-97 listed in section 4.6.1.3, which is a different ICP method than that listed for Fe (I-1472-87) in Table 4. K needs a separate analytical description if it is being analyzed by flame AA as stated in the method cited. (Also a description in the calibration section for K analysis)	reference to methods id number deleted from section 4.6.1.3. Method ID listed in table 4
VAC23	What about other method required QC : Serial dilutions or interference check stds?	A complete description of QC checks is listed for method I-4471-97 is described in Garbarino, J.R., and Struzeski, T.M., 1998, Methods of analysis by the U.S. Geological Survey National Water Quality Laboratory -- Determination of elements in whole-water digests using inductively coupled plasma- optical emission spectrometry and inductively coupled plasma-mass spectrometry: U.S. Geological Survey Open-File Report 98-165, 101 p. QC information generated in the analytical process is retained by the laboratory and available on request.	40	Complete (Table 4)	
VAC24	Micro-related sections are currently out to our Microbiologist at the lab; awaiting comments on procedures and method.	Comments related to bacteria analysis listed below microNU1-microNU6			

JC25	What is the criteria for the blanks? How will blank results be evaluated? What corrective action or data validation will occur if they are outside of the criteria?	Laboratory blank must be less than the long-term method detection limit (LT-MDL); if analysis of blank samples is greater than LT-MDL affected samples will be rerun. Field blanks will be evaluated for sampling contamination, if value exceeds two times the long-term detection limit or is within 10 percent of the mean sample concentration. samples will be flagged as estimated values due blank contamination and efforts will be made to identify and eliminate the source of contamination.	40	Same criteria for Filtration Blank and Equipment Blank? (Currently the criteria cited is listed under Field blank only)	Criteria the same for all blank samples. Text page 38
VAC26	Needs QC table for lab analyses with acceptance criteria by analysis for the QC listed in this section. (Blanks, MS/MSD, dup, surrogates, etc). While the lab has their determined QC criteria, it needs to be stated in the QAPP what the project goals are so it is a stand-alone document.	nutrients, regresion eqn plus/minus 1.5 fpsuedo sigma all sample values must be bracketed by QA data within control limits.		?? Not sure if I got your complete comment, but I think this is satisfied by the generic criteria supplied in Table 3 - Complete	ok
VAC27	What is released, i.e. what level of deliverables will the lab be providing? If 'levels' are not defined, state in detail what the lab will be providing: data result reports and an analysis narrative? Raw data?	Proprietary data is released to the NWIS database and WaWSC pending final review by project staff	43	Complete (5.0) I was looking for the type of deliverables released by the lab, but it sounds like it is only the final results, no lab QC.	Data from the Blind Standard Reference sample programe is continual released as seperated standalong data base. Bench QA and continuing calibration data retained, (eventuallyarchived) and available on request. Page 44
VAC28	Who applies data qualifiers? Will any lab qualification occur? What qualifiers are used/definition. U, J, R etc	Data qualifiers can be applieied either at the lab or by project/review personnel.	40	Complete (4.9.2)	
JC29	What about data sharing with WCD and EPA for the entire ARM project? State when / how the data will be provided to other parties and specifically who the contacts are that would be receiving the data. EPA/USGS expectations for data sharing is probably found in the interagency agreement and may be appropriate to state/reference here as well.	Data sharing between USGS and WCD will be o continious process conducted by individual project chiefs or their designates. Logistical details of this data sharing will be disscussed and documented at the initiaition of field sampling.	44	Complete (5.1)	
VAC 30	Please reference EPA G5/G4 for QAPP guidance and DQO development	noted and done	10	Complete (references)	

microNU1	Make sure that the samples collected for fecal coliform are collected aseptically and that the other testing mentioned as field screening is not done on just a portion of the pump sample. Preferably, the sample should be collected first for the coliform testing. Will they use an EPA certified lab for the testing? How will they clean or sanitize the sampling device between samples assuming they collect from more than one site during an event? Peristaltic pumps make it easy to just change out the entire tubing with new sterile tubing – hopefully that is their intent.	Aseptic techniques will be used for all micor sampling and equipment and buffer blanks are included as part of all bacteria sampling runs. Much of the micro field techniques are described in chapter 7 of USGS Field Manual which includes such items as not rinsing sample bottle, use of sodium thiosulfate to neutralize bleach used to field sterilize.	32	Complete (4.3.1)	
microNU2	Need to be more specific – the hold time is actually 8 hours for anything that is not drinking water. However, if they wanted to use the 24 hour hold time, they should specify this rather than saying 1 day.	Hold time is 8 hours, although I think our (USGS) guidance is 6hr.	36	Please update in Table 4 from 1 day to the HT which will be adhered to in this project. WCD was allowed 24 hours due to storm events/etc but they are going to try their best to meet the EPA prescribed HT (ECY allows 24 hours)	24 hours used in table 4
microNU3	Doesn't work for microbiology. They should not field rinse the bottle and the bottle should be sterile – hence no field rinsing. PE is usually sterilized using irradiation or gas as it doesn't tolerate the pressure/heat associated with autoclaves. They don't identify the "C" in RUC in this table.. does that mean chilled?	I believe the sample bottles we autoclave are constructed of HDPE. Could sterile Whirl pac bags be used as sample containers for groundwater and wastewater sample collection.	36	I have no clue about Whirl pacs - if I am thinking of the correct baggie, I have seen it used for soils but not waters. You can clarify for me!	
microNU4	This could be a big problem unless they ensure that all the chlorine residual is removed from the tubing prior to sample collection. They could neutralize the chlorine by flushing the line with sodium thiosulfate... or just water and then testing the water for chlorine prior to sample collection for bacteria.	sodium thiosulfate rinse is part of the protocol	32	Complete	
microNU5	All good stuff. Especially if they make sure that the tubing used for collection is free of chlorine prior to sample collection.	Can check rinse solution with chloine test strips. H	36	Complete. A check rinse for chlorine with test strips sounds like a good idea to verify the tubing is free of chlorine prior to micro collection.	page 36
microNU6	There will be a difference in results between USGS (E. coli) and Whatcom's fecal coliform testing. Usually (but not always) fecal coliform counts will be higher.....	This is one of the discussion point that are scheduled to be hammered out between WCD and USGS in the early phase of field sampling so that comparability of data is maximized.	36	Complete - noted in QAPP and above.	

	Steve, Here is some language for the criteria for deviating from the target of 4 wells on each parcel and the clear statement that the intention is to install 4 unless some serious technical or agricultural challenge drives you to drop to 3...Since 2 wouldn't allow us to figure out even the flow direction, I just can't consider 2 a reasonable number for this project...				
Curt1		Language was changed to reflect the intent to install 4 wells per plot area.	10	Looks like it was addressed to me - please verify with Curt	see page10
Curt2	Same language as above and rationale	Language was changed to reflect the intent to install 4 wells per plot area.	10	Looks like it was addressed to me - please verify with Curt	
Kozar1	my only major concern is related to the use of packers in the screened interval of the 2-inch wells. I know that you are also somewhat concerned about the potential for cross-contamination within the filter pack of the well.	Use of a very fine grained sand, much finer than the aquifer material to be sampled, will be used in the annular space around the screened portion of the well to mitigate any potential vertical flow from one packed interval to the next.	21		
Kozar2	Check performance of the multiple-zone packer assembly to isolate sampling zones.	injection of a fluoromteric tracer in the interval below the lower most packer and sampling of the overlying packed intervals for presence to the tracer will help to verify that the packer assembly is working as designed, and that cross contamination between packers is not occurring or is minimal.	21		
Kozar3	minimize the potential for inducing a head change over the multiple-packed intervals.	The low pumping rate (roughly 10 ml/min) should minimize the potential for induced head gradients between sampling intervals.	24		

Assessment of variability in analytical concentrations

Sequential replicates	Variability related to sample collection, processing and short term local variability.
Split replicates	Variability related to analytical process
Blank	Identify sample bias/contamination